

## Original Article

# *Nauclea Latifolia* (African Peach) Attenuates Gastric Acid Secretion in Rats with Ethanol-Induced Ulceration

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### ABSTRACT

**Objective:** The ethanolic leaf extract of *N. latifolia* (EENL) was investigated for its gastric acid antisecretory effects on ethanol-induced gastric ulcerated rats using an *in vivo* assay. **Methods:** Gastric mucosal ulceration was induced experimentally in Groups 2 to 5 rats using ethanol. Five groups of rats were orally pretreated with 3% Tween-80 only as vehicle group, 3% Tween-80 as ulcer group, 20 mg/kg of omeprazole as positive group, 250 and 500 mg/kg of EENL as the test groups, respectively before ulcer induction. Gastric acid secretion parameters and ulcerated areas were evaluated. **Results:** Pretreatment with EENL showed significant ( $P < 0.05$ ) reduction in gastric volume, total acidity and ulcer index while the pH of gastric content increased significantly ( $P < 0.05$ ) in rats with ethanol-induced ulcer. Ulcer group exhibited significantly severe mucosal injury and EENL or omeprazole offered significant ( $P < 0.05$ ) protection against gastric mucosal injuries. **Conclusion:** The study suggests that EENL attenuates gastric acid secretion and ulceration induced by ethanol in Wistar rat. The observed pharmacological activities might be mediated via antisecretory and cytoprotective mechanisms.

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### Introduction

Ulcer is a major disease of gastrointestinal system which accounts for estimated 15 mortalities out of 15,000 complications annually (Milivojevic and Milosavljevic, 2020). Even though scientists and medical researchers cannot fully ascertain the exact pathogenesis of gastric ulcer, a common ground has been established. Gastric mucosal ulceration occurred whenever the stomach secretion of aggressive factors

(acid and pepsin) overwhelm that of protective factors (mucus and bicarbonate) resulting in disruption of the gastric mucosal layer (Malik *et al.*, 2018). The pathogenesis of gastric ulcer is multifactorial in nature, these include: increased acid-pepsin secretion (Silva and de Sousa 2011), impaired bicarbonate neutralization and reduced mucus secretion (Balogun *et al.*, 2018b). A great number of available orthodox drugs (such as proton pump inhibitors, histamine

receptor antagonists and antibiotics) have been employed in treatment and management of gastric mucosal ulceration. Nevertheless, the major challenges remain the adverse effects and resistance of these drugs with prolonged use (Alaribe *et al.*, 2014). These triggered the quest for safer and cheaper antiulcer natural drugs. In view of the above challenges, it is pertinent to investigate the phytotherapy and effectiveness of medicinal plant formulations employed in folkloric medicine for treatment, management and prevention of gastric ulcer. Numerous medicinal plants used in animal experiments have been documented to possess antiulcer activities (Balogun *et al.*, 2015; Iheagwam *et al.*, 2020). *Nauclea latifolia* Smith (*N. latifolia*), commonly known as African peach tree, is an emerging plant of interest in the treatment, prevention and management of gastric ulcer. It is a flowering plant belonging to the family of *Rubiaceae*, and genus of *Sarcocephalus*. It is commonly known as pin cushion tree being a straggling shrub or small tree, native to the tropical Africa and Asia (Udobi *et al.*, 2017).

It is known locally as “Ubulu inu” among the Igbo in the eastern part of Nigeria; as “Tafashiya” among the Hausas in the northern Nigeria; as “Egbesi” by Yorubas in the western part of Nigeria and as “Itu” among the Itsekiri (Arise *et al.*, 2012; Balogun *et al.*, 2016a). The leaves of *N. latifolia* have many therapeutic applications in folk medicine due to their effective secondary metabolites such as flavonoids, cardiac glycosides, saponin and tannin (Onyesom *et al.*, 2015; Iheagwam *et al.*, 2020). The sticks are used as chewing stick and a remedy against tuberculosis (Balogun *et al.*, 2013; Nyong *et al.*, 2021). Several scientific studies have been documented on its biological potencies such as hypolipidemic and hypoglycemic, antiulcer, antihypertensive, antioxidative and hepatoprotective effects in laboratory animal experiments (Balogun *et al.*, 2015; Onyesom *et al.*, 2015; Oyetayo *et al.*, 2020; Busari *et al.*, 2021). In spite of that, experimental evidence in favour of gastric acid antisecretory potential of ethanolic leaf extract of *N. latifolia* is yet to be reported. Therefore, the current study aimed to investigate the gastric acid antisecretory effects of the ethanolic leaf extract of *N. latifolia* (EENL) on ethanol-induced ulcerated rats.

## Materials and Methods

### Drug and reagents

All chemicals, drugs, and reagents used in this study were obtained commercially and of analytical grade. Tween 80, ethanol, diethyl ether, HCl and NaOH were procured from Sigma-Aldrich Chemical Company (St. Louis Missouri, USA). Distilled water was obtained

from Physiology Laboratory, Al-Hikmah University, Ilorin, Nigeria.

### Drug preparation

Omeprazole (Globela Pharma PVT. Ltd., India), procured from One Step Pharmacy, Ilorin, Nigeria, was used as standard antiulcer drug in this study. The drug was dissolved in 3% Tween-80 and administered to the animals orally in a dose of 20 mg/kg body weight (5 mL/kg) prior to gastric ulcer induction as earlier described by Balogun *et al.*, (2018a).

### Experimental animals

Adult male (8-10 weeks old) Wistar rats weighing ( $172.9 \pm 6.03$ ) g were used for the study. The rats were obtained from the Faculty of Health Sciences, Central Animal House, Al-Hikmah University, Ilorin. They were kept under standard laboratory conditions and fed with commercial grower's feed (Vital feeds Nig Ltd, Nigeria) and water *ad libitum*. The animals were allowed one-week acclimatization before the experiment commenced.

### Ethical Approval

The research was conducted following the approval by the ethics committee for animal experimentation of the Faculty of Health Sciences, Al-Hikmah University, Ilorin (Ethic No. HUI/REC/FHS/2021/02) and animal handling was according to accepted guidelines by the National Institutes of Health for care and use of laboratory animals (NRC, 2010).

### Harvesting and authentication of plant material

Fresh leaves of *N. latifolia* were collected from a farm settlement in Iseyin Local Government Area of Oyo State, Nigeria. The leaves were identified and authenticated by Mr. K.A. Adeniji of the Forestry Research Institute of Nigeria (FRIN) Ibadan, Nigeria. A voucher specimen (FHI. 110284) was deposited in the herbarium of the institute for future references.

### Preparation of leaf extract

Freshly collected leaves were thoroughly washed with distilled water and air dried at room temperature. The leaf powder (1200 g) was extracted with absolute ethanol (96%, v/v) in a soxhlet extractor at 70°C. The mixture was vacuum-filtered through Whatman No. 1 filter paper and the filtered extract was concentrated using a vacuum rotary evaporator (Eyla N-1000, Japan) maintained at 45°C. The resulting residue which weighed 98.6 g (recovery 8.2%) was later stored under 4°C before use. Prior to oral administration of the extract, the required dose of 250 or 500 mg/kg body weight was obtained after the extract was reconstituted in 3% Tween-80 (Balogun *et al.*, 2018b; Iheagwam *et al.*, 2020).

**Preliminary phytochemical analysis**

The EENL was subjected to qualitative phytochemical screening to pinpoint the presence of secondary constituents (such as alkaloids, saponins, terpenoids, glycosides, anthraquinones, flavonoids, tannins, resins, cardiac glycosides, steroids and phenols) using standard phytochemical methods described by Harborne, (1988).

**Acute oral toxicity studies**

The “fixed dose” method of Organization for Economic Cooperation and Development (2008) guideline No. 425 was employed to determine the acute oral toxicity of EENL in adult albino rats (OECD, 2008). The method was commenced with an initial dose of 2000 mg/kg body weight after overnight dietary deprivation. The animals were observed for general behavioural, autonomic and neurological behaviour during the course of the experiment.

**Experimental design and treatment**

Gastric ulcer was induced experimentally in groups 2 to 5 using ethanol model. The rats were allotted into five groups of five rats in each group. Group 1 (normal control) received 3% Tween-80 only. Group 2 (ulcer control) received 3% Tween-80. Group 3 (standard) received 20 mg/kg omeprazole while groups 4 and 5 (experimental groups) received 250 and 500 mg/kg body weight (5 mL/kg body weight) of EENL respectively. The administration was done by oral gavage for all groups prior to ulcer induction.

**Ethanol-induced gastric ulcer**

Gastric mucosal ulceration was induced experimentally using ethanol-induced model previously described by Mizui and Douteuchi (1983). In this method, gastric lesion was induced in 24 h fasted rats by the oral administration of 1 mL/kg of ethanol (80% ethanol) 30 min after 3% Tween-80, omeprazole (20 mg/kg body weight), EENL (250 and 500 mg/kg body weight) were administered.

**Excision of stomach and gastric juice collection**

The rats were sacrificed humanely under diethyl ether (Sigma-Aldrich, USA) anaesthetization after 1 h of ethanol administration. The stomach was excised and opened along the greater curvature following the opening of the abdomen. The gastric content of the stomach was drained into a centrifuge tube in addition with 5 mL of distilled water. The resultant solution was centrifuged at  $3000 \times g$  for 10 min at 4 °C and the supernatant collected was subjected to biochemical analyses thereafter (Shay *et al.*, 1954).

**Ulcer score and percentage inhibition**

The gastric mucosal layer of the stomach was viewed under a magnifying lens (10x) to evaluate the gastric lesions. The number of ulcers were counted and scored using the method described by Kulkarni *et al.*, (1987). The score was graded as 0, normal colouration, 0.5, red colouration; 1.0, spot ulcers; 1.5, haemorrhagic streaks; 2.0, deep ulcers; 3.0, perforations. The sum of ulcer scores assigned to gastric lesions was expressed as the mean ulcer index (MUI) (Tan *et al.*, 1996). Percentage of ulcer inhibition was calculated according to Hojage *et al.* (2010) using the formula: Percentage inhibition (%I) =  $(MUI_{\text{control}} - MUI_{\text{test}}) \div MUI_{\text{control}} \times 100\%$

**Determination of gastric juice volume and pH**

The supernatant fluid volume was measured in millilitre (mL) using micro syringe (Bickel and Herling, 2002). The gastric content samples were analysed for hydrogen ion concentration using a digital pH meter (Abdulla *et al.*, 2010).

**Determination of total acidity and gastric acid output**

One millilitre of gastric juice was titrated with 0.01N (mEq/L) NaOH in a conical flask using phenolphthalein (two drops) as indicator until light pink solution indicating pH 7.0 was obtained Wang *et al.*, (2007). The volume of NaOH added was employed in the calculation to get the total acidity using the formula below (Raju *et al.*, 2009).

Total acidity =  $\text{Volume of NaOH} \times \text{normality of NaOH} \times 100 \text{ (mEq/L)} \div 0.1$

Acid output was expressed as micro equivalents per hour ( $\mu\text{Eq/hr}$ ) and calculated by multiplying the total acidity in mEq/L by the volume of gastric juice in (litre) (Brodie and Hook, 1971). The result was divided by 4 to give output per hour.

Acid output ( $\mu\text{Eq/hr}$ ) =  $\text{Acidity (mEq/L)} \times \text{Volume of gastric juice (ml)} \div 4 \text{ hr}$ .

**Gross evaluation of gastric mucosa**

The mucosal layer of the stomach of each rat was rinsed with normal saline to remove blood clot if any. The stomach was then pinned to a flat board to observe any changes in the physical appearance of the mucosa. Photographs of the gastric lesions were taken for proper observation and documentation.

**Statistical analysis**

The values were expressed as mean  $\pm$  standard error of mean. For data comparison, one-way analysis of variance (ANOVA) was used followed by Tukey's multiple comparison tests. Differences between groups were considered statistically significant at  $P < 0.05$  using Graph pad Prism Version 6.0 for Windows (GraphPad® Software, San Diego, CA, USA).

## Results

### Extract yield

The ethanolic extraction of 1200 g of *N. latifolia* leaf powder yielded 8.2% (w/w) greenish semisolid extract with a pasty consistency.

### Qualitative phytochemical analysis

The result of the preliminary qualitative phytochemical studies of EENL showed the presence of saponins, tannins, cardiac glycosides, alkaloids, flavonoids, terpenoids and phenols.

### Acute oral toxicity studies

All the animals remained alive and showed no visible signs of toxicity even with the highest dose. There were no abnormal signs, changes in body weight and behaviour in the EENL-treated animals throughout the

observation period when compared to normal control animals. Thus, the median lethal dose was considered to be greater than 2000 mg/kg body weight.

### Effect of EENL on ulcer index and ulcer inhibition

The effects of EENL on ulcer index and inhibition are reported in Table 1. Ulcer index in the ulcer control group was significantly ( $P < 0.05$ ) increased as compared to compared vehicle control group. The severity of ethanol-induced gastric ulcer was significantly ( $P < 0.05$ ) reduced by pretreatment with EENL or omeprazole. The extract exhibited a dose-dependent inhibition against ulceration in rats. However, maximum inhibition was observed in omeprazole pretreated group, which was similar to the 500 mg/kg EENL.

**Table 1:** Effects of EENL on ulcer index and ulcer inhibition in ethanol-induced ulcerated rats

Group	Treatment and dose	Ulcer index	Ulcer inhibition (%)
1 (Vehicle control)	Tween 80 (2 mL/kg)	0.00 ± 0.00	100.00
2 (Ulcer control)	Tween 80 (2 mL/kg) + ethanol (1 mL/kg)	13.60 ± 0.40 <sup>a</sup>	0.00
3 (Standard)	Omeprazole (20 mg/kg) + ethanol (1 mL/kg)	1.30 ± 0.82 <sup>b</sup>	90.4
4 (Test)	EENL (250 mg/kg) + ethanol (1 mL/kg)	2.00 ± 0.77 <sup>b</sup>	85.3
5 (Test)	EENL (500 mL/kg) + ethanol (1 mL/kg)	1.50 ± 0.59 <sup>b</sup>	89.0

All values are expressed as mean ± standard error of mean (n = 5 in each group). <sup>a</sup> $P < 0.05$ , vs normal control group; <sup>b</sup> $P < 0.05$ , vs ulcer control group. EENL: ethanolic leaf extract of *Nauclea latifolia*.

### Effect of EENL on gastric juice volume and pH

The effects of EENL on gastric juice volume and pH are reported in Table 2. The gastric juice volume was significantly increased ( $P < 0.05$ ) in the ulcer control group with concomitant decrease in pH when compared to the vehicle control group. The group pretreated with 250 mg/kg or 500mg/kg EENL produced a significant ( $P < 0.05$ ) and dose-dependent

( $P < 0.05$ ) decrease in gastric juice volume with corresponding increase in pH compared to the ulcer control group. Meanwhile, a significant ( $P < 0.05$ ) decrease in gastric volume with concomitant increase in pH was observed in groups pretreated with omeprazole (20 mg/kg) in comparison to the ulcer control group.

**Table 2:** Effects of EENL on gastric juice volume and pH in ethanol-induced ulcerated rats

Group	Treatment and dose	Gastric content (mL)	pH
1 (Vehicle control)	Tween 80 (2 mL/kg)	7.43 ± 2.94	4.03 ± 3.95
2 (Ulcer control)	Tween 80 (2 mL/kg) + ethanol (1 mL/kg)	13.49 ± 2.87 <sup>*</sup>	1.92 ± 1.88 <sup>*</sup>
3 (Standard)	Omeprazole (20 mg/kg) + ethanol (1 mL/kg)	5.31 ± 3.62 <sup>#</sup>	6.80 ± 2.03 <sup>#</sup>
4 (Test)	EENL (250 mg/kg) + ethanol (1 mL/kg)	6.05 ± 2.83 <sup>#</sup>	5.99 ± 4.67 <sup>#</sup>
5 (Test)	EENL (500 mL/kg) + ethanol (1 mL/kg)	5.70 ± 4.39 <sup>#</sup>	6.47 ± 5.43 <sup>#</sup>

All values are expressed as mean ± standard error of mean (n = 5 in each group). <sup>\*</sup> $P < 0.05$ , vs vehicle control group; <sup>#</sup> $P < 0.05$ , vs ulcer control group; EENL: ethanolic leaf extract of *Nauclea latifolia*.

### Effect of EENL on total acidity and gastric acid output

The effects of EENL on total acidity and gastric acid output are shown in Table 3. Total acidity and acid output were significantly increased ( $P < 0.05$ ) in the ulcer control group compared with vehicle control

group. Omeprazole produced a significant decrease ( $P < 0.05$ ) in total acidity and acid output as compared with the ulcer control. A significant ( $P < 0.05$ ) and dose dependent ( $P < 0.05$ ) decrease in total acidity and acid output were observed in the EENL-pretreated groups in comparison to ulcer control group.

**Table 3:** Effects of EENL on total acidity and gastric acid output in ethanol-induced ulcerated rats

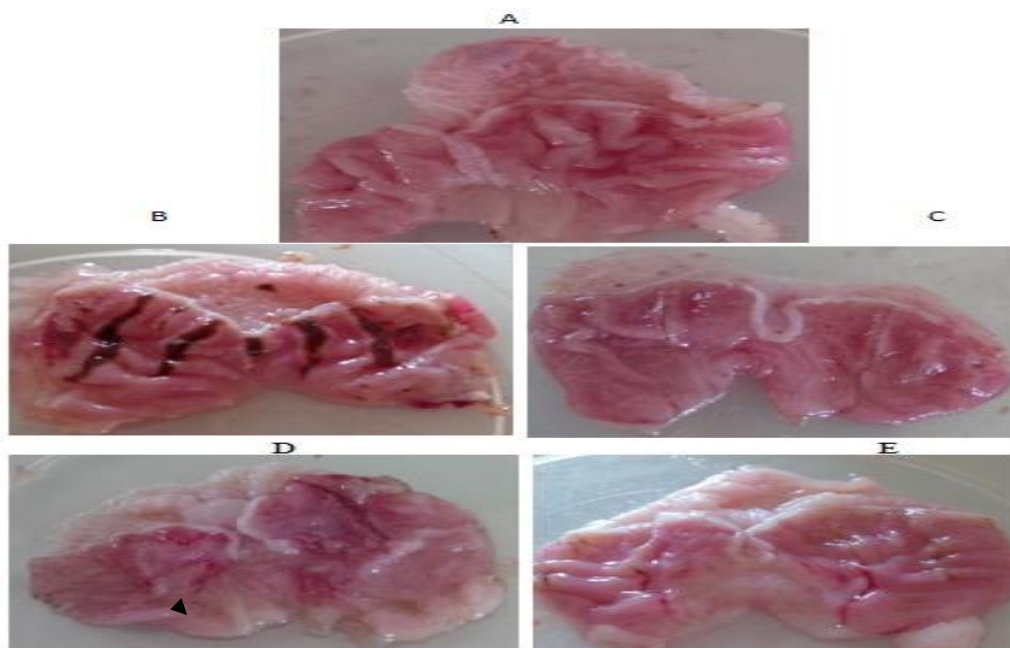
Group	Treatment and dose	Total acidity (mEq/L)	Acid output ( $\mu$ Eq/h)
1 (Vehicle control)	Tween 80 (2 mL/kg)	62.38 $\pm$ 1.93	34.63 $\pm$ 3.06
2 (Ulcer control)	Tween 80 (2 mL/kg) + ethanol (1 mL/kg)	89.73 $\pm$ 1.69*	67.09 $\pm$ 5.43*
3 (Standard)	Omeprazole (20 mg/kg) + ethanol (1 mL/kg)	48.39 $\pm$ 1.37 <sup>#</sup>	30.54 $\pm$ 6.11 <sup>#</sup>
4 (Test)	EENL (250 mg/kg) + ethanol (1 mL/kg)	57.05 $\pm$ 1.50 <sup>#</sup>	43.66 $\pm$ 4.08 <sup>#</sup>
5 (Test)	EENL (500 mg/kg) + ethanol (1 mL/kg)	52.84 $\pm$ 1.76 <sup>#</sup>	36.17 $\pm$ 5.56 <sup>#</sup>

All values are expressed as mean  $\pm$  standard error of mean (n = 5 in each group). \* $P < 0.05$ , vs vehicle control group; <sup>#</sup> $P < 0.05$ , vs ulcer control group; EENL: ethanolic leaf extract of *Nauclea latifolia*.

#### Effects of EENL on macroscopic appearance of the gastric mucosa

In Fig. 1, the vehicle control group showed normal gastric mucosal architecture. Ulcer control group showed severe gastric mucosal injuries with extensive

haemorrhagic necrosis of the gastric mucosa. EENL-pretreated groups (250 and 500 mg/kg) exhibited a fairly protected mucosa in ethanol-induced ulcer model compared to the injuries seen in the ulcer control rats.



**Fig. 1:** Gross appearance of the gastric mucosa in ethanol-induced ulcerated rats. A: Vehicle control group; B: Ulcer control group; C: Omeprazole group; D: EENL (250mg/kg) group; E: EENL (500mg/kg) group. EENL: ethanolic leaf extract of *Nauclea latifolia*.

#### Discussion

There is increasing attention on the management and prevention of gastric ulcer through identification and evaluation of drugs from natural products as an alternative treatment approach. The present study employed ethanol-induced ulcer model to investigate the possible gastric acid antisecretory effects EENL. Based on the available research findings, the study has shown that EENL possessed significant gastric acid antisecretory activity on ethanol-induced ulcerated rats. Our results show that EENL is capable of attenuating ethanol-induced gastric mucosal

ulceration in rats. The extract at low and high doses offered 85.3% and 89.0% mucosal protection respectively against ethanol-induced ulcerations. Interestingly, the extract at both doses, attenuated an increase in gastric acid secretion and mucosal injuries similar to omeprazole. This is an indication that extract competes favourably with the reference drug. However, the mechanisms behind these gastric acid antisecretory effects of EENL in rats have not yet been investigated. The pathogenic effects of ethanol-induced lesions have been established to include increased secretion of gastric acid, which resulted in

increased gastric volume, reduced pH, and increased total acidity, acid output and ulcer index (Zayachkivska *et al.*, 2005). The auto digestion of the mucosal wall by accumulated gastric juice causes mucosal barrier breakdown leading to gastric ulcer formation (Vinothapooshan *et al.*, 2010). Findings from this study show that in ethanol-induced ulcer model, the EENL (250 and 500 mg/kg) pretreated groups produced a significant decrease in acid output, total acidity, and volume of gastric content with a corresponding increase in pH compared to the ulcer control group. This decrease in gastric juice volume in the extract pretreated groups may be due to a decrease in acid production as evidenced from the gastric juice total acidity. This is an indication that EENL contains some biological compounds that reduce the acidity of gastric secretions which was increased by ethanol. Agents that inhibit the effects of ethanol will exhibit cytoprotection. The results from this study showed that EENL can significantly prevent the gastric acid secretory effects of ethanol by exhibiting a fairly protected mucosa with a significant decrease in ulcer index and gastric acid production in comparison to the ulcer control group. This observed cytoprotection offered by the extract may be mediated by strengthening of the mucosal barrier through increase in prostaglandin synthesis enabling it to resist the toxic effects of ethanol. Ethanolic leaf extract of *N. latifolia* has been shown to contain phytochemicals such as flavonoids, cardiac glycosides and terpenoids which are among the cytoprotective materials known to possess antiulcer (Balogun *et al.*, 2018b), antioxidant (Iheagwam *et al.* 2020) activities respectively. The active compounds, flavonoids and cardiac glycosides which are present in the leaves have also been reported by Onyesom *et al.*, (2015) and Iheagwam *et al.* (2020) as antioxidant materials. Therefore, it is pertinent that the decrease in gastric lesions, decrease in volume and acidity of gastric fluid and decrease in gastric acid production by EENL in this study could be due to the presence of these active components or some other mechanisms yet to be unravelled. In conclusion, the study suggests that EENL possesses significant gastroprotective effects against ethanol-induced gastric ulcer in rats. The observed gastric acid antisecretory effects might due to its antisecretory and cytoprotective properties. If this is applied to man, our findings may have beneficial application in the management of gastric mucosal lesions associated with ethanol-induced gastric ulceration.

#### Competing interests

The authors declare no conflict of interest.

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